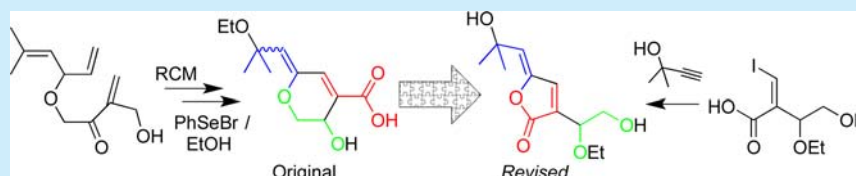


Total Synthesis and Structural Revision of the Cytotoxin Aruncin B

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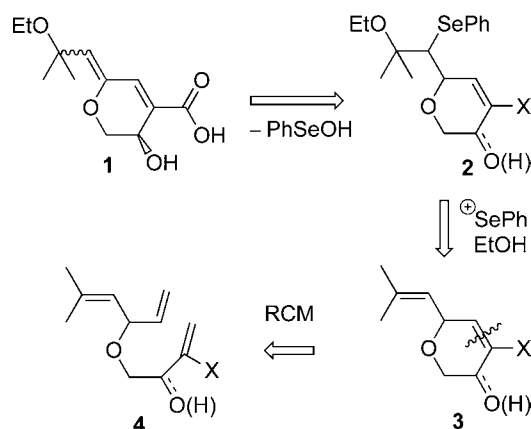
S Supporting Information



ABSTRACT: The sodium salts *E*-15 and *Z*-15 of the originally proposed dihydropyran acid structure of aruncin B (**1**) were prepared through ring-closing alkene metathesis (RCM) and ethoxyselenation–selenoxide elimination, but acid sensitivity of these salts, together with inconsistencies in the spectral data, suggested a significant structural misassignment. A β -iodo Morita–Baylis–Hillman reaction to give *Z*-iodo ester **24**, followed by Sonogashira cross-coupling–5-*exo-dig* lactonization, provided concise access to a Z - γ -alkylidenebutenolide **18**, which possessed data corresponding to those originally reported for aruncin B.

In 2011, Woo and co-workers reported cytotoxicity-directed isolation of extracts from the plant *Aruncus dioicus* var. *kamtschaticus*, obtained from the Korean island Ulleungdo, which enabled discovery of the monoterpenoid (+)-aruncin B (**1**) (Scheme 1, 29 mg from 6.9 kg of aerial parts).¹ Studies indicated

Scheme 1. Structure of Aruncin B (1**) Assigned by Woo and Co-workers,¹ and Retrosynthetic Analysis**



inhibition of Bcl-2 antiapoptotic proteins, with selective cytotoxicity for malignant tumor cells over normal human T cells suggesting potential as an antitumor agent.^{1b}

The structure of aruncin B was assigned using spectroscopic methods (NMR, MS, UV, and IR) and shows interesting features: an enol ether with a unique cross-conjugated pattern, along with a potentially acid-labile tertiary allylic ethyl ether coexisting with carboxylic acid functionality. The exocyclic double bond geometry and absolute configuration were not established. The intriguing structure and biological activity make aruncin B (**1**) an attractive target for synthesis. Late-stage installation of the potentially sensitive enol ether motif was

envisaged using an ethoxyselenation–selenoxide elimination sequence² (Scheme 1, **3** \rightarrow **2** \rightarrow **1**). The precursor dihydropyran-containing core **3** would be formed by ring-closing alkene metathesis (RCM) of a triene **4**. Appropriate ring and side-chain (X) oxidation levels for this strategy would be established through experimentation.

The synthesis of the postulated structure began with 1,2-addition of vinylmagnesium bromide to prenal (**5**) (Scheme 2), followed by *O*-alkylation of the resulting crude dienol³ with *tert*-butyl bromoacetate, under phase transfer conditions,⁴ to give ester **6**. Following phosphonation,⁵ phosphonate **7** underwent the Villieras variant of the Horner–Wadsworth–Emmons reaction⁶ to give, after purification by column chromatography (the first required in this sequence), α -hydroxymethyl enone **8**. RCM⁷ of the latter using Grubbs' II catalyst gave hydroxymethyldihydropyranone **9**. Due to metathetical involvement of the *gem*-dimethyl substituted alkene, ~10% of undesired dihydropyranone **10** was originally obtained as an inseparable RCM byproduct; this issue was overcome by subsequent addition of excess 2-methyl-2-butene.⁸

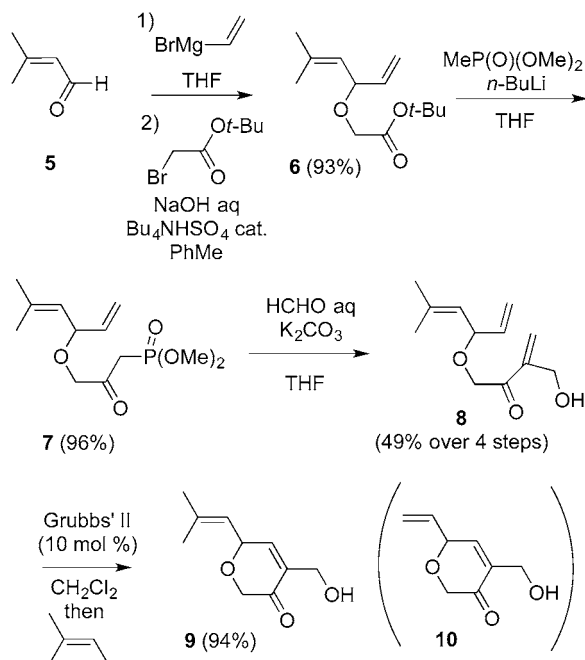
Hydroxymethyldihydropyranone **9** contains all the core carbon atoms of aruncin B. Reduction using DIBALH⁹ gave a diol **11** (Scheme 3, 3:1 dr) from which the primary alcohol was selectively oxidized to the hydroxyaldehyde **12** through TEMPO catalysis;¹⁰ no ketone was observed. For analytical convenience the hydroxyaldehyde **12** diastereoisomers were separated at this stage, and subsequent chemistry was carried out on the major (*trans*¹¹) diastereoisomer.

Ethoxyselenation² occurred selectively at the electron-rich olefin of the hydroxyaldehyde **12mj**, to give the ethoxyselenide **13** as a single diastereoisomer (Scheme 4).¹² Periodate oxidation¹³ of the crude ethoxyselenide **13** followed by

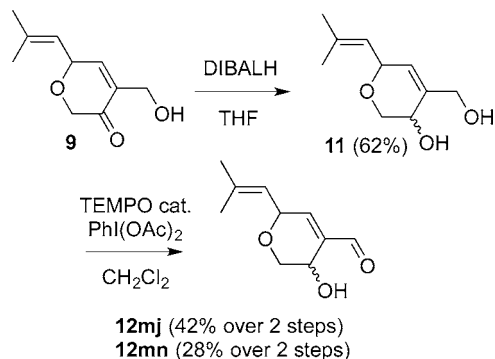
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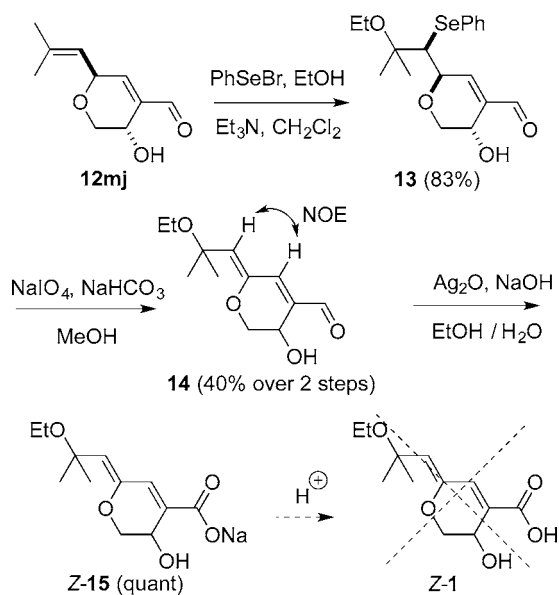
Scheme 2. Synthesis of Hydroxymethyldihydropyranone 9



Scheme 3. Redox Adjustment

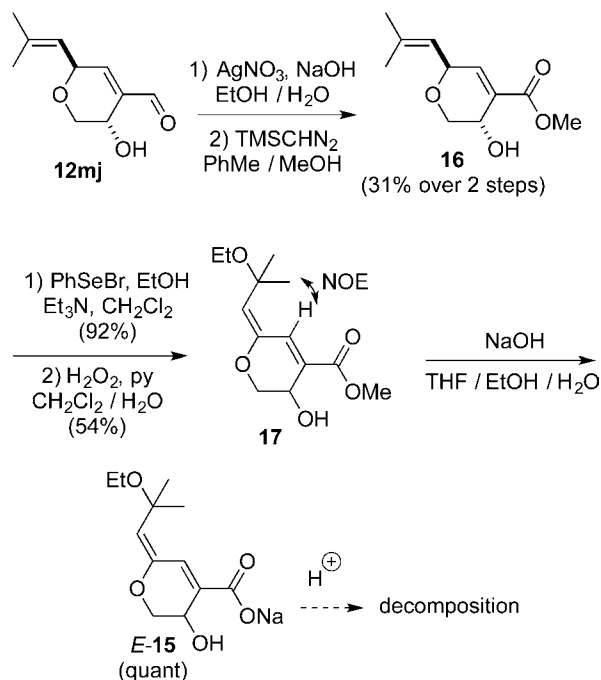


Scheme 4. Ethoxyselenation–Selenoxide Elimination Sequence, Final Oxidation and Attempted Acidification



selenoxide elimination gave the desired enol ether **14** as the *Z*-isomer. The *E*-isomer was observed in the crude mixture¹⁴ but not isolated; it is suspected to be the initial product from *syn* elimination, but isomerization of the double bond occurs rapidly (possibly by reversible conjugate addition at the δ -position of the dienal) to deliver the *Z*-isomer, likely favored due to minimization of allylic strain. Final chemoselective oxidation of the aldehyde to the carboxylic acid was anticipated to be achievable using freshly prepared silver oxide under alkaline conditions.¹⁵ While this proceeded smoothly to deliver the Na-salt **Z-15** of the originally assigned structure, all attempts to obtain the free acid from **Z-15** led only to complete and rapid decomposition. For example, mild acidification conditions, such as pH 6 buffer solution, addition of accurately measured 1 equiv of HCl (at $-78\text{ }^{\circ}\text{C}$), or carboxylic acid-supported ion-exchange resins all proved unsuccessful.

Since enol ether geometry of aruncin B had not been established in the isolation paper,^{1a} we were interested in seeing whether the *E*-isomer would possess different acid stability character and correspond to the natural product. It was considered that switching from an aldehyde to a slightly less electron-withdrawing group, such as an ester, might retard isomerization and thereby enable the access to the *E*-configured exocyclic unsaturation. Oxidation¹⁶ of hydroxyaldehyde **12mj**, then esterification using TMSCHN₂, delivered hydroxy ester **16** (Scheme 5). Ethoxyselenation as before, followed by hydrogen

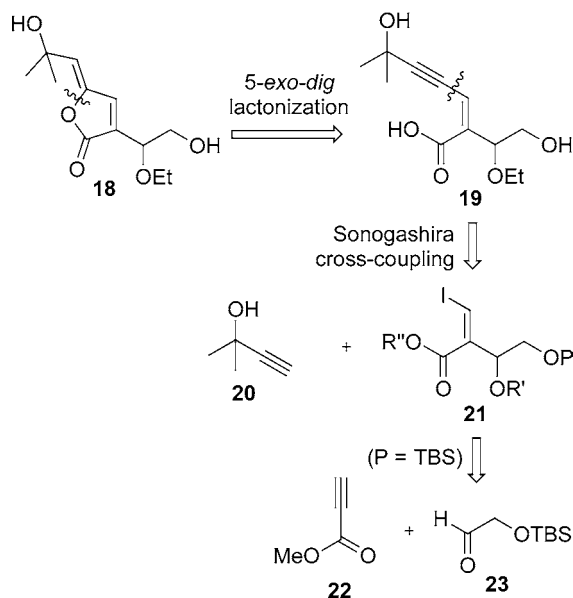
Scheme 5. Synthesis of the Na-Salt *E*-15

peroxide-induced oxidation¹⁷ of the resulting selenide, gave the *E*-enol ether **17**, which exhibited greater configurational stability than enol ether **14**. Saponification of *E*-enol ether **17** delivered the *E*-Na-salt **E-15** which, however, possessed the same sensitivity to acid as previously found with *Z*-Na-salt **Z-15**.

Our above studies suggested that acid **1** is intrinsically unstable, due to the coexistence of tertiary allylic ether and carboxylic acid functionality: acid-induced loss of EtOH, assisted by the enol ether, leads to decomposition. While UV data supported the presence of a γ -alkoxy- $\alpha,\beta,\gamma,\delta$ -unsaturated

carboxyl motif,^{18,19} acid sensitivity, together with inconsistencies in the ¹H and ¹³C NMR data between **1** and the two Na-salts (**Z-15** and **E-15**),²⁰ indicated that a structural revision was necessary. Detailed analysis of the data suggested that, in particular, a better match for the originally assigned ring CH₂ would be as a primary alcohol,²¹ rather than an (enol) ether. Further inspection of the original structure assignment of aruncin B (**1**)^{1a} indicated that the site of etherification was not determined unambiguously and, based on the ¹³C NMR data, a structure featuring tertiary allylic alcohol and the ethyl ether of a secondary alcohol would be a more reasonable alternative.²² These considerations led us to consider a *Z*- γ -alkylidenebutenolide **18** (Scheme 6) as being

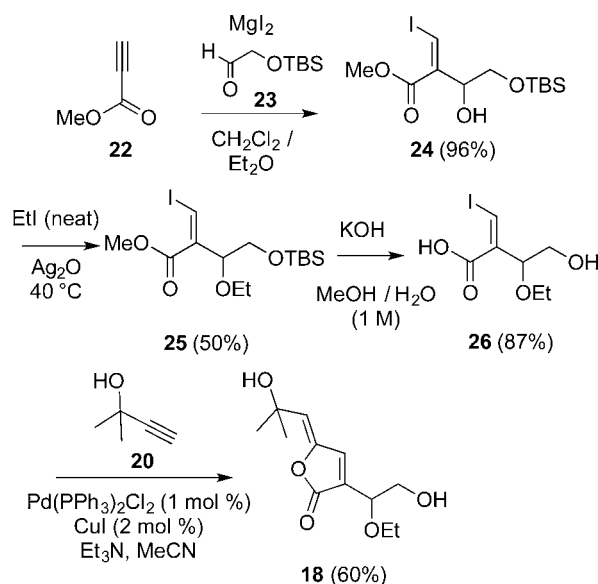
Scheme 6. New Postulated Structure of Aruncin B (Butenolide 18) and Retrosynthetic Plan



more consistent with available data.²⁴ The γ -alkylidenebutenolide motif is present in several natural products,²⁵ including novaxenicin D²⁶ and an oxidized tirucallane triterpenoid²⁷ which possess *E*- γ -(hydroxy)isobutylidene and *Z*- γ -(methoxy)isobutylidene substitution, respectively. However, the α -dioxxygenated side chain has not previously been observed. While many routes to γ -alkylidenebutenolides are known,²⁵ we were attracted to a Sonogashira cross-coupling–5-*exo-dig* lactonization sequence²⁸ (**20** + **21** \rightarrow **19** \rightarrow **18**, Scheme 6) due to its functional group tolerance, stereocontrol, and brevity. The *Z*-vinyl iodide substrate **21** bearing the requisite α -dioxxygenated side chain could be prepared via a β -iodo-Morita–Baylis–Hillman (MBH) reaction.²⁹

The synthesis starts with methyl propiolate (**22**) and the aldehyde **23**.³⁰ The β -iodo-MBH reaction was carried out using *in situ* generated MgI₂ as an iodide source,³¹ giving *Z*-iodoester **24** (Scheme 7). This powerful one-step transformation installs the dioxxygenated side chain and sets the *Z*-vinyl iodide functionality necessary for the subsequent cross-coupling. *O*-Ethylation of the secondary alcohol was achieved using Ag₂O in neat EtI, to give the ether **25**. Subsequent saponification of the methyl ester was accompanied by desilylation to give the desired cross-coupling substrate *Z*-iodoacid **26**. The Sonogashira cross-coupling–5-*exo-dig* lactonization sequence could be achieved in the presence of free alcohol functionality in both partners, and with as little as 1 mol % Pd(PPh₃)₂Cl₂ and 2 mol % CuI (~1

Scheme 7. Synthesis of the Correct Structure of Aruncin B: Butenolide 18

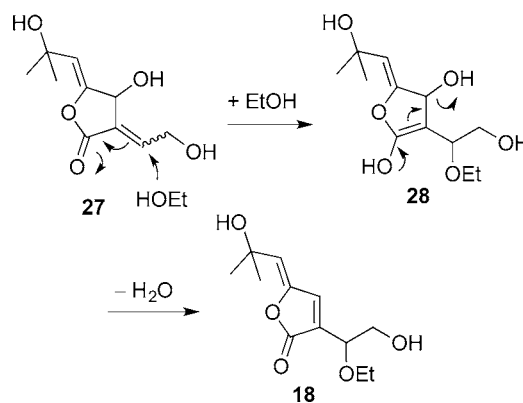


mmol scale, leading to 200 mg of *Z*-butenolide **18**). To our delight, the spectroscopic data for synthetic γ -alkylidenebutenolide **18** corresponded to those reported for the originally isolated aruncin B.^{20,32}

Enantiopure samples of (+)- and (–)-aruncin B were obtained from *Z*-butenolide **18** by semiprep HPLC. The specific rotation values found for the synthetic materials ($[\alpha]_D^{25}$ +/– 90 (*c* 1.0 in MeOH)) were significantly higher than that reported for the natural isolate ($[\alpha]_D^{20}$ + 27 (*c* 0.1 in MeOH)).^{1b} This discrepancy suggests that the original isolate may not be enantiopure, but rather partially enantioenriched. An intriguing structural feature of the correct structure of aruncin B is the ethoxy group at the β' -position. Interestingly, aruncin B was isolated by EtOH extraction of the aerial plant parts. The possibility of low enantiomeric enrichment and unusual presence of the ethoxy group could indicate that butenolide **18** is an artifact of the isolation process, resulting from a modestly diastereoselective conjugate addition of EtOH to an α -(2-hydroxyethylidene)- γ -lactone **27**,³³ followed by loss of water (Scheme 8).

In summary, RCM followed by ethoxyselenation–selenoxide elimination proved to be a viable strategy to (ethoxy)-isobutylidene-substituted dihydropyransols, which was applied in stereocontrolled syntheses of the Na-salts of the *Z*- and *E*-

Scheme 8. Potential Origin of Butenolide 18



isomers of the originally postulated structure of aruncin B (**1**). However, the free acids could not be obtained from these Na-salts. This instability, together with inconsistencies in the collected analytical data, led to the conclusion that aruncin B has been misidentified.³⁴ An alternative butenolide **18** structure has been proposed and synthesized in four steps, through β -iodo-MBH reaction and Sonogashira cross-coupling–lactonization. The data for butenolide **18** match with those originally reported for aruncin B. With the structure of aruncin B now reassigned and a concise and flexible synthetic approach to it established, studies can now be undertaken to probe in more detail the biological activity of aruncin B and analogues.

■ ASSOCIATED CONTENT

■ Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: [10.1021/acs.orglett.6b02120](https://doi.org/10.1021/acs.orglett.6b02120).

Experimental procedures, characterization data, and copies of ^1H and ^{13}C NMR spectra for all reported compounds (PDF)

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Notes

The authors declare no competing financial interest.

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■ REFERENCES

- (1) (a) Jeong, S. Y.; Jun, D. Y.; Kim, Y. H.; Min, B.-S.; Min, B. K.; Woo, M. H. *Bioorg. Med. Chem. Lett.* **2011**, *21*, 3252–3256. (b) Han, C. R.; Jun, D. Y.; Woo, H. J.; Jeong, S.-Y.; Woo, M.-H.; Kim, Y. H. *Bioorg. Med. Chem. Lett.* **2012**, *22*, 945–953.
- (2) (a) Nicolaou, K. C.; Magolda, R. L.; Sipio, W. J.; Barnette, W. E.; Lysenko, Z.; Joullie, M. M. *J. Am. Chem. Soc.* **1980**, *102*, 3784–3793. (b) Garratt, D. G.; Kabo, A. *Can. J. Chem.* **1980**, *58*, 1030–1041. (c) Foote, K. M.; Hayes, C. J.; Pattenden, G. *Tetrahedron Lett.* **1996**, *37*, 275–278.
- (3) Austin, K. A. B.; Elsworth, J. D.; Banwell, M. G.; Willis, A. C. *Org. Biomol. Chem.* **2010**, *8*, 751–754.
- (4) Yip, K.-T.; Li, J.-H.; Lee, O.-Y.; Yang, D. *Org. Lett.* **2005**, *7*, 5717–5719.
- (5) Sauerland, S. J. K.; Castillo-Meléndez, J. A.; Nättinen, K.; Rissanen, K.; Koskinen, A. M. P. *Synthesis* **2010**, 2010, 757–762.
- (6) (a) Villieras, J.; Rambaud, M. *Synthesis* **1983**, 1983, 300–303. (b) Paterson, I.; Tillyer, R. D.; Ryan, G. R. *Tetrahedron Lett.* **1993**, *34*, 4389–4392. (c) Ryan, S. J.; Thompson, C. D.; Lupton, D. W. *Aust. J. Chem.* **2009**, *62*, 720–727.
- (7) Taillier, C.; Hameury, T.; Bellosta, V.; Cossy, J. *Tetrahedron* **2007**, *63*, 4472–4490.
- (8) Chatterjee, A. K.; Sanders, D. P.; Grubbs, R. H. *Org. Lett.* **2002**, *4*, 1939–1942.
- (9) Li, C.; Porco, J. A., Jr. *J. Org. Chem.* **2005**, *70*, 6053–6065.
- (10) (a) Lin, S.; Dudley, G. B.; Tan, D. S.; Danishefsky, S. J. *Angew. Chem., Int. Ed.* **2002**, *41*, 2188–2191. (b) Mandal, M.; Yun, H.; Dudley, G. B.; Lin, S.; Tan, D. S.; Danishefsky, S. J. *J. Org. Chem.* **2005**, *70*, 10619–10637.
- (11) Assigned according to ring $^3J_{\text{H-H}}$ values; see the [Supporting Information](#).
- (12) Acidic workup was used to deprotect small amounts of the corresponding diethyl acetal of **13** formed during the course of the reaction.
- (13) Sharpless, K. B.; Lauer, R. F.; Teranishi, A. Y. *J. Am. Chem. Soc.* **1973**, *95*, 6137–6139. Peroxide-based methods were avoided, due to potential Baeyer–Villiger chemistry at the aldehyde.
- (14) Characteristic ^1H NMR olefinic shifts for the *E*-isomer are 7.93 and 5.46 ppm. A rationale for why the *E*-isomer might be initially favored is given in the [Supporting Information](#).
- (15) (a) Su, Z.; Tamm, C. *Helv. Chim. Acta* **1995**, *78*, 1278–1290. (b) Pearl, I. A. *Org. Synth.* **1963**, Coll. Vol. 4, 972–976.
- (16) Cambie, C.; Denny, W. A.; Hay, M. P.; Mitchell, L. H.; Rutledge, P. S.; Woodgate, P. D. *Aust. J. Chem.* **1999**, *52*, 7–18.
- (17) Mastalerz, H.; Menard, M.; Vinet, V.; Desiderio, J.; Fung-Tomc, J.; Kessler, R.; Tsai, Y. J. *Med. Chem.* **1988**, *31*, 1190–1196.
- (18) Silverstein, R. M.; Clayton, G.; Bassler, B.; Morrill, T. C. *Spectrometric Identification of Organic Compounds*, 5th ed.; Wiley: New York, 1991; pp 301–304.
- (19) Woodward, R. B.; Weiler, L. S.; Dutta, P. C. *J. Am. Chem. Soc.* **1965**, *87*, 4662–4663.
- (20) See the [Supporting Information](#) for tabular NMR comparison.
- (21) Tori, M.; Takeichi, Y.; Kuga, H.; Nakashima, K.; Sono, M. *Chem. Pharm. Bull.* **2002**, *50*, 1250–1254.
- (22) (a) Vassilikogiannakis, G.; Stratakis, M.; Orfanopoulos, M.; Foote, C. S. *J. Org. Chem.* **1999**, *64*, 4130–4139. (b) Pei, Y.-G.; Wu, Q.-X.; Shi, Y.-P. *J. Chin. Chem. Soc.* **2007**, *54*, 1565–1572. (c) Reisch, J.; Herath, H. M. T. B.; Bergenthal, D.; Kumar, N. S. *Liebigs Ann. Chem.* **1991**, 1991, 1233–1235.
- (23) Findlay, J. A.; Li, G.; Miller, J. D.; Womiloju, T. O. *Can. J. Chem.* **2003**, *81*, 284–292.
- (24) While the anticipated C=O IR data for **18** ($\sim 1755\text{ cm}^{-1}$, cf., ref 26) would not be consistent with those reported for **1** (1724 cm^{-1} , KBr disc, ref 1), the original IR spectrum could not be located (Prof. Woo, personal communication ninth July 2015) and none of the natural isolate was available for further analysis.
- (25) For reviews on γ -alkylidenebutenolides, see: (a) Negishi, E.; Kotora, M. *Tetrahedron* **1997**, *53*, 6707–6738. (b) Brückner, R. *Curr. Org. Chem.* **2001**, *5*, 679–718. (c) Barbosa, L. C. A.; Teixeira, R. R.; Pinheiro, P. F.; Maltha, C. R. A.; Demuner, A. J. *Quim. Nova* **2010**, *33*, 1163–1174. (d) Barbosa, L. C. A.; Teixeira, R. R.; Amarante, G. W. *Curr. Org. Synth.* **2015**, *12*, 746–771.
- (26) Bishara, A.; Rudi, A.; Goldberg, I.; Benayahu, Y.; Kashman, Y. *Tetrahedron* **2006**, *62*, 12092–12097.
- (27) Wang, J.-S.; Zhang, Y.; Wei, D.-D.; Wang, X.-B.; Luo, J.; Kong, L.-Y. *Chem. Biodiversity* **2011**, *8*, 2025–2034.
- (28) Lu, X.; Huang, X.; Ma, S. *Tetrahedron Lett.* **1993**, *34*, 5963–5966.
- (29) Taniguchi, M.; Hino, T.; Kishi, Y. *Tetrahedron Lett.* **1986**, *27*, 4767–4770.
- (30) Commercially available (e.g., Acros, Aldrich, Fluorochem), or prepared in two steps from 2-butene-1,4-diol: Vanier, S. F.; Larouche, G.; Wurz, R. P.; Charette, A. B. *Org. Lett.* **2010**, *12*, 672–675.
- (31) Matsuda, Y.; Kato, M.; Kawaguchi, T.; Koyama, T.; Saikawa, Y.; Nakata, M. *Tetrahedron* **2014**, *70*, 1154–1168.
- (32) Excepting the C=O IR data for γ -alkylidenebutenolide **18** (1755 cm^{-1} , KBr disc), which was as expected.
- (33) Many monoterpenoid α -(2-hydroxyethylidene)butanolides are known; for example: (a) Ma, L.-J.; Wang, Y.-H.; Tang, G.-H.; Wang, Y.-L.; Ma, C.; Dastmalchi, K.; Kennelly, E. J.; Long, C.-L. *Planta Med.* **2013**, *79*, 308–311. (b) Yoshida, K.; Hishida, A.; Iida, O.; Hosokawa, K.; Kawabata, J. *J. Nat. Prod.* **2010**, *73*, 814–817.
- (34) For a review on natural product misassignment, see: Nicolaou, K. C.; Snyder, S. A. *Angew. Chem., Int. Ed.* **2005**, *44*, 1012–1044.